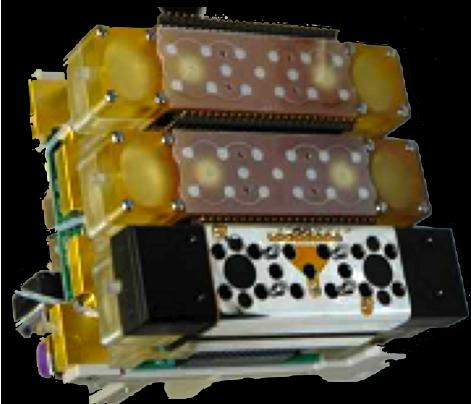
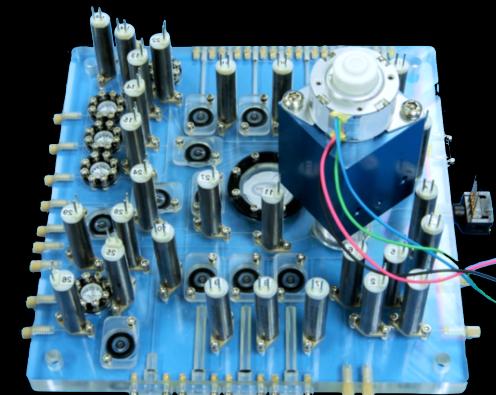




Leveraging Cubesat Payload Technologies to Search for Life on our Solar System's Icy Worlds



Tony Ricco
NASA Ames Research Center



with many thanks to
ARC small payloads and search-for-life tech. dev. teams
and the space biologists & astrobiologists everywhere who
Define the science that defines the tech



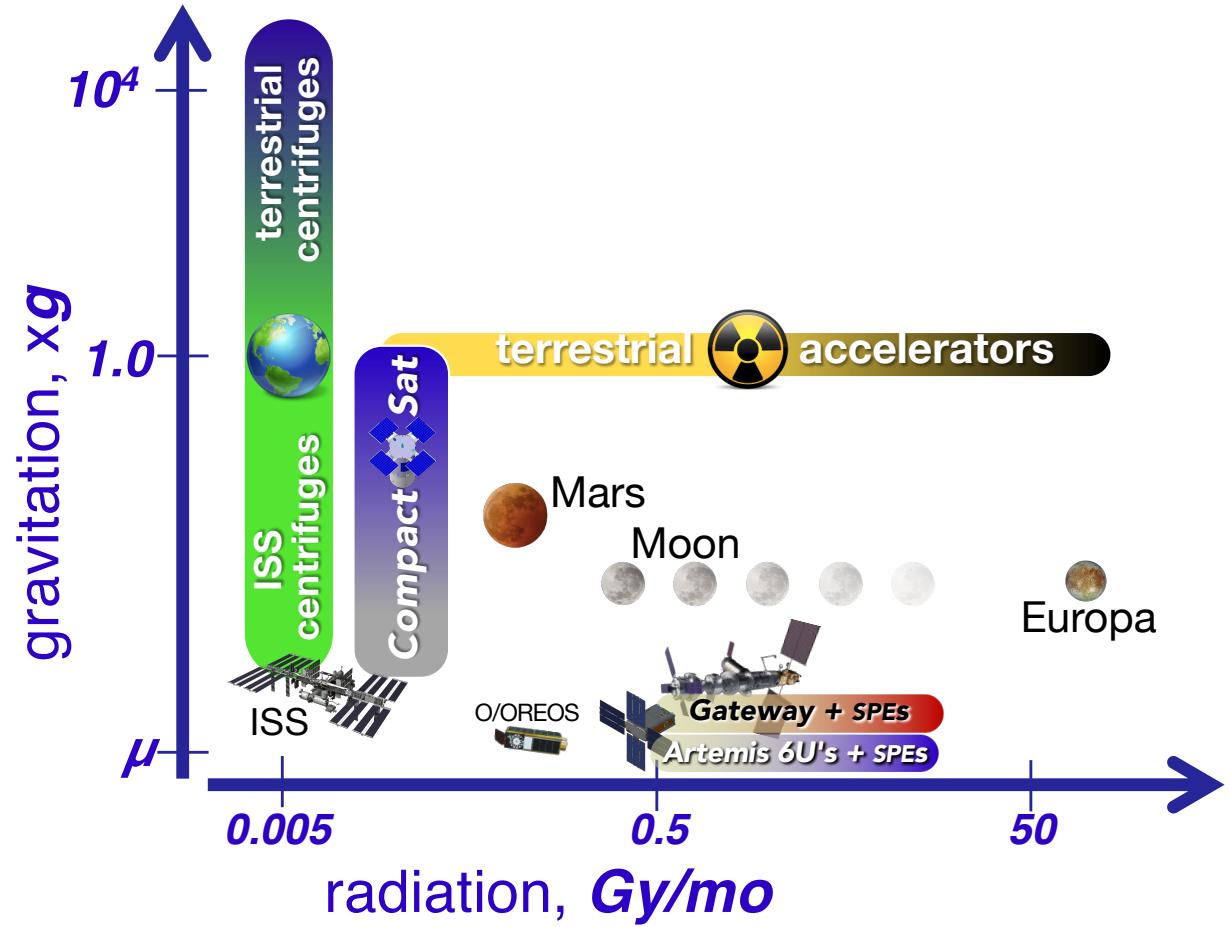
National Aeronautics and
Space Administration



“BioMapping” the solar system and its gravity-radiation landscape:

- where can terrestrial life exist/thrive?
- where does, or did, non-terrestrial life exist?

*Further considerations:
temperature, pressure, chemistry*

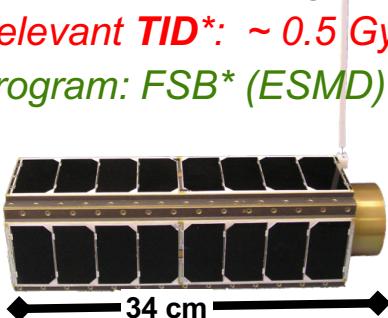




Integrated Microfluidic Bioanalytical Systems to Grow and Monitor Microbes in Space

GeneSat (2004-2006-2010)

- Orbit: Low Earth Orbit, 440 km
- Mission duration: 1 month
- Orbital lifetime: 3.7 years
- Relevant TID*: ~ 0.5 Gy
- Program: FSB* (ESMD)

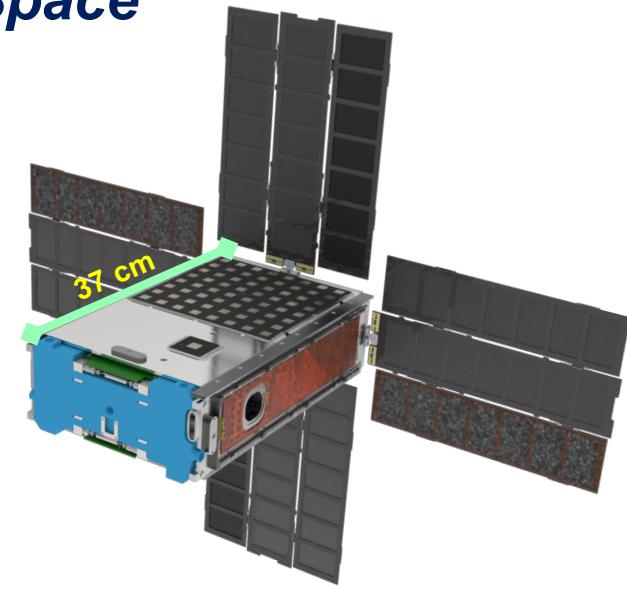
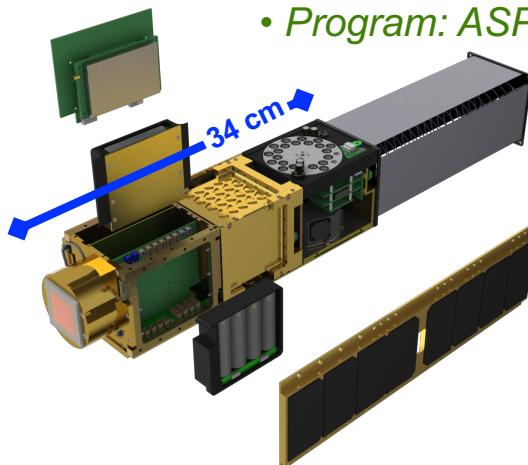


*TID = total ionizing dose

*FSB = Fundamental Space Biology

O/OREOS (2008-2010-2032)

- Orbit: High-inclination LEO, 650 km
- Mission duration: 6 – 18 months
 - Orbital lifetime: ~22 years
 - Relevant TID*: 1 – 50 Gy
 - Program: ASP* (SMD)

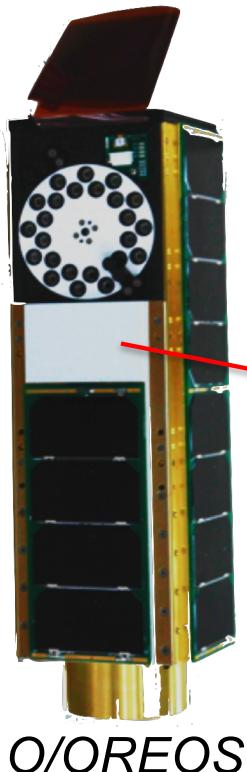


BioSentinel (2013-2021-7500000000)

- Orbit: Interplanetary (heliocentric), 100 k – 60 M km
- Mission duration: 3 – 9 months
- Orbital lifetime: ~ 7.5×10^9 years
- Relevant TID*: ~ 3 Gy
- Program: AES* (HEOMD)

*ASP = Astrobiology Small Payloads *AES = Advanced Exploration Systems

Summary of NASA Ames' Nanosatellite (Astro)Biological Space Missions



O/OREOS



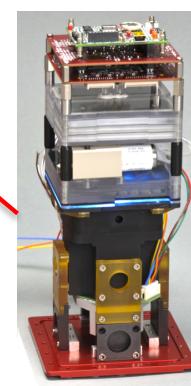
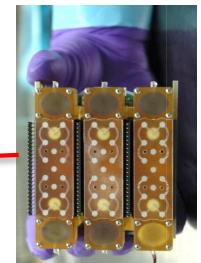
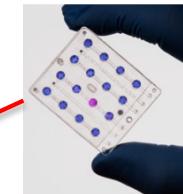
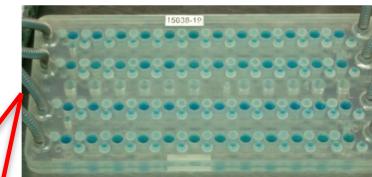
E. Coli GeneSat-1 (2006/3U): *gene expression*
EcAMSat (2017/6U): *antibiotic resistance*

S. Cerevisiae PharmaSat (2009/3U): *drug dose response*
BioSentinel (2021/6U): *DNA break/repair*

B. Subtilis O/OREOS* (2010/3U): *survival, metabolism*
ADRoIT-M** (6U): *mutations / lithopanspermia*

Ceratopteris Richardii SporeSat-1 (2014/3U): *ion channel sensors, µ-centrifuges*
SporeSat-2 (3U): *plant gravity sensing threshold*

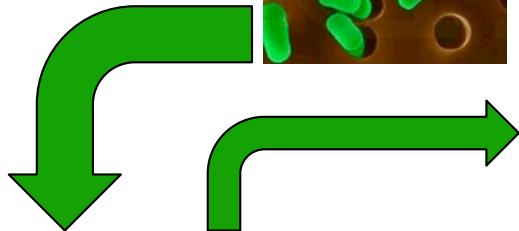
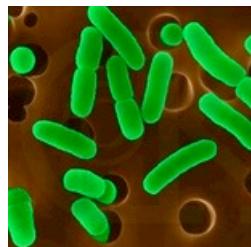
C. Elegans FLAIR (3U):
dual-wavelength fluorescence imager



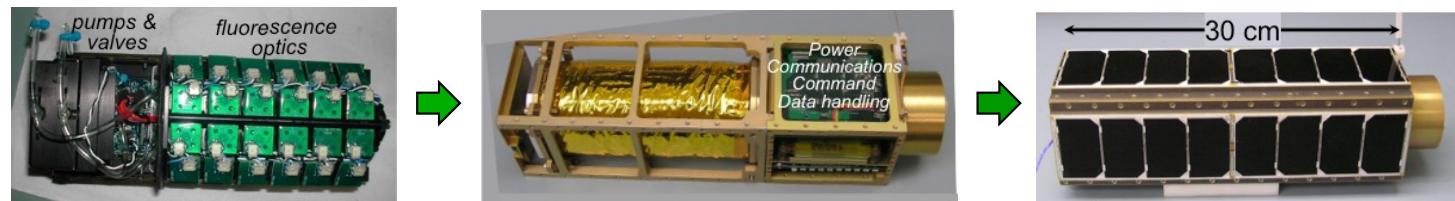
*Organism/Organic Exposure to Orbital Stresses

**Active DNA Repair on Interplanetary Transport of Microbes

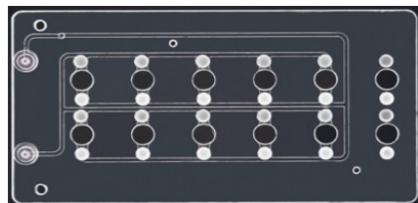
model organism:
0.5 x 2 μ m
bacteria
E. coli



GeneSat-1: 1st biological nanosatellite in LEO, 1st real-time, gene expression measurement in space



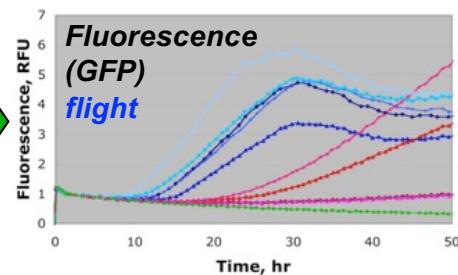
- nutrient deprivation in dormant state (6 weeks)
- launch: December 2006 to low Earth orbit (440 km)
- nutrient solution feed upon orbit stabilization, grow *E. coli* in μ gravity
- monitor green fluorescent protein (**GFP**): gene expression
- monitor optical density: cell population



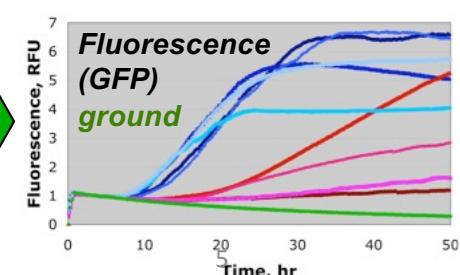
16 December
2006



Telemeter data
to Earth



Compare to
ground data



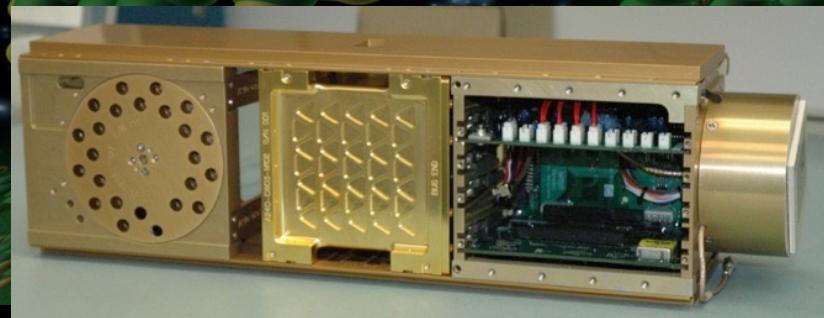
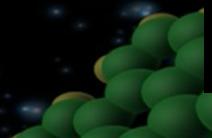


Kodiak bear

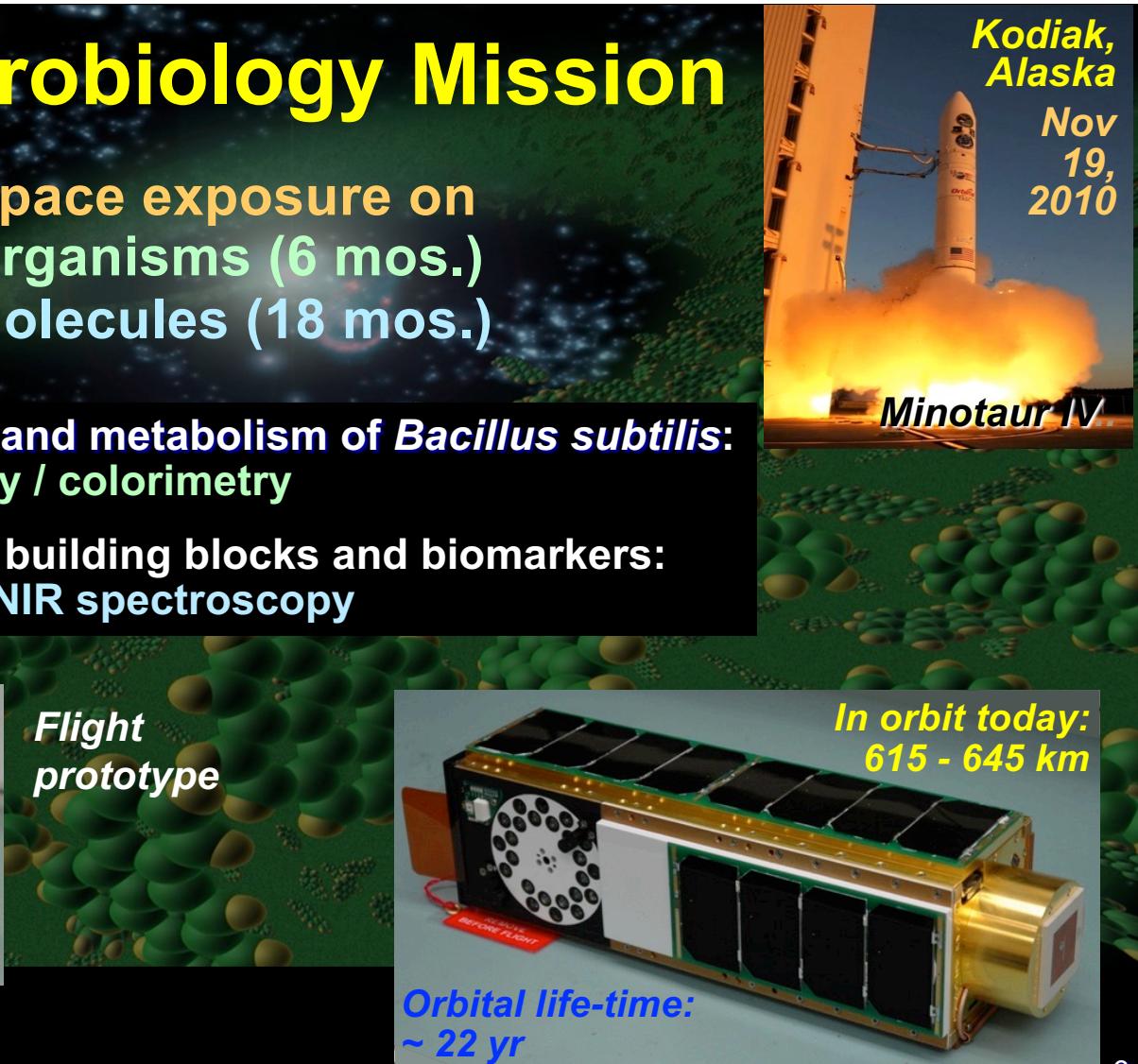
O/OREOS Astrobiology Mission

Effects of space exposure on
biological organisms (6 mos.)
& organic molecules (18 mos.)

- Monitor survival, growth, and metabolism of *Bacillus subtilis*:
in-situ optical density / colorimetry
- Track changes in organic building blocks and biomarkers:
UV / visible / NIR spectroscopy



Flight
prototype



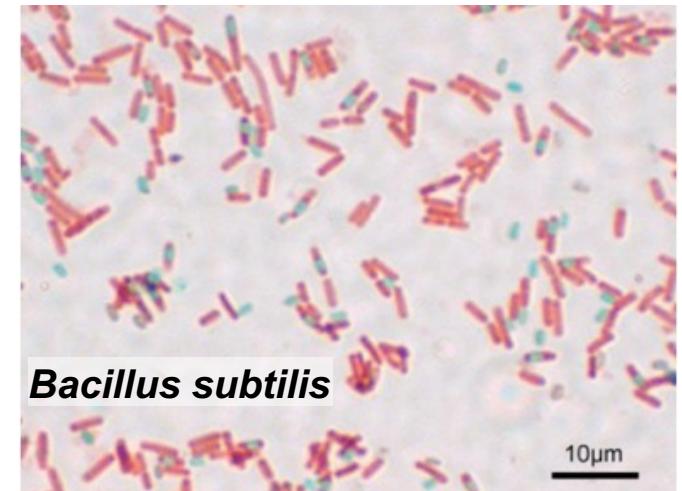
Ehrenfreund *et al*, Acta Astro 93, 501 (2014)



Payload 1: Space Environment Survivability of Live Organisms (SESLO)

Organisms, wildtype & mutant, exposed to μ gravity & space radiation

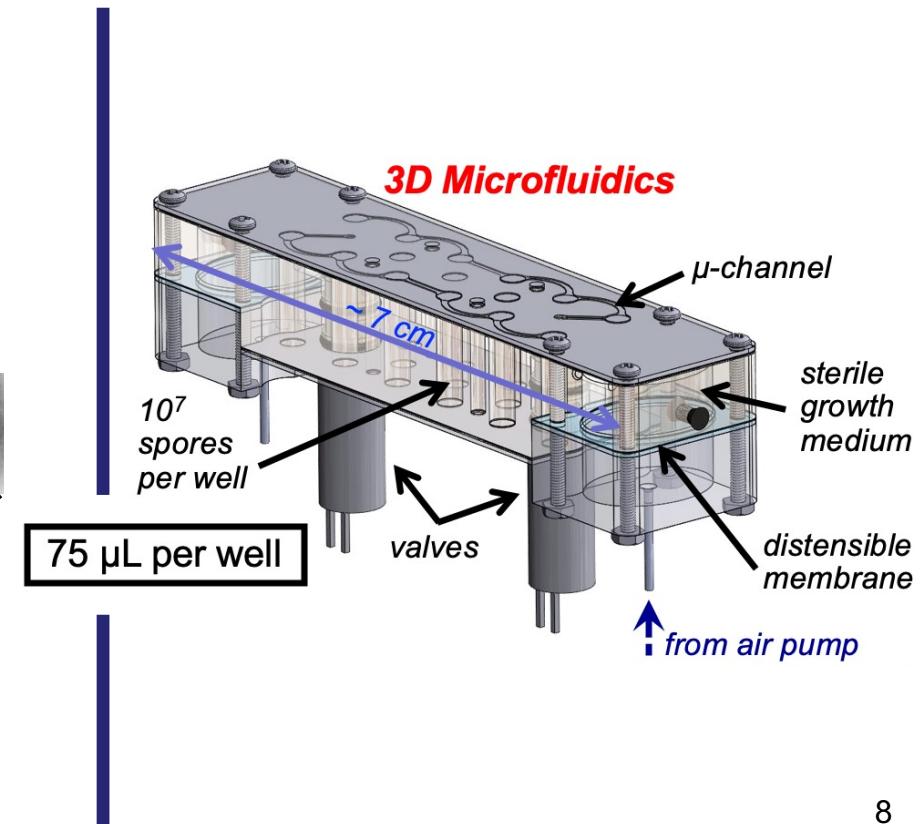
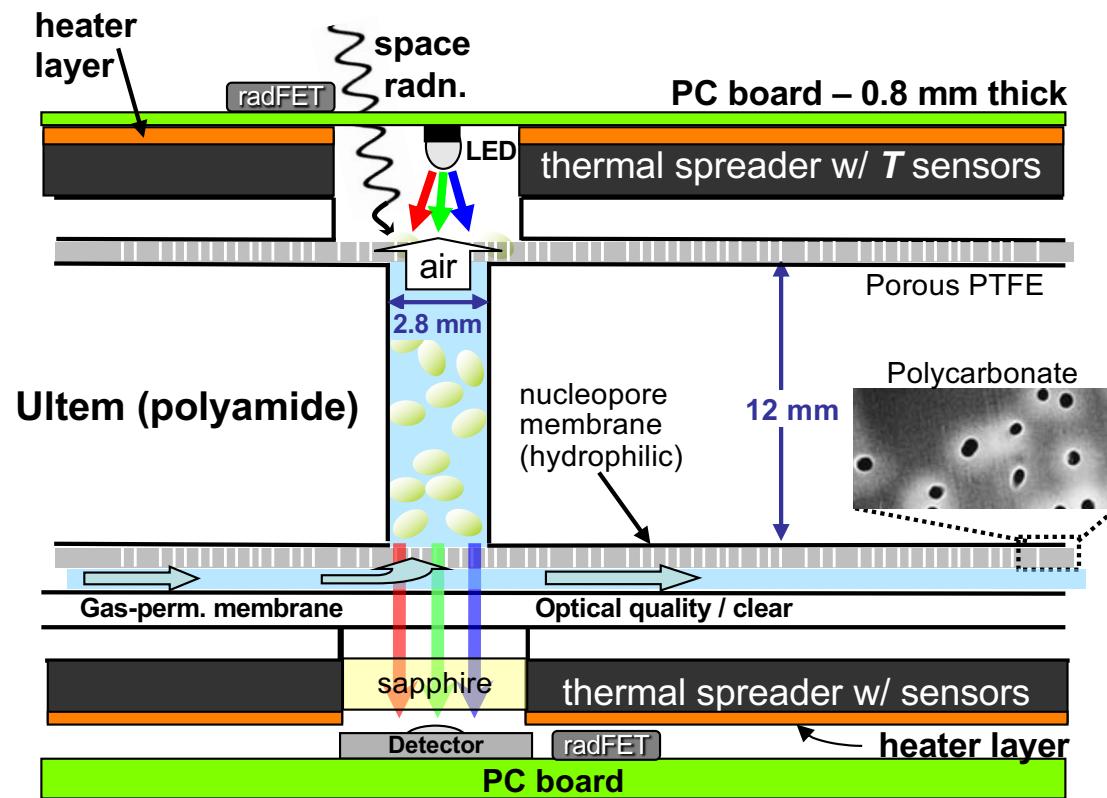
- On Earth: dry organisms on microwell walls
- In space: Rehydrate & feed 6 μ wells / organism:
 $t = 2 \text{ wk}, 3 \text{ mo}, 6 \text{ mo}$ (**requires perfect sterility**)
- Grow @ **37 °C** for ~3 days
- Measure **RGB absorbance@ 615, 525, 470 nm**
 - track culture population via optical density
 - track metabolic activity via [**Alamar Blue**]
- Sensors: **T, p, RH, rad** (integrated dose), **μ grav**
 - » temperature (6 sensors per 12-well bioblock)
 - » pressure, relative humidity (1 sensor each)
 - » radiation total dose @ both ends of wells (2 radFETs)
 - » microgravity levels calc'd. from solar panel currents

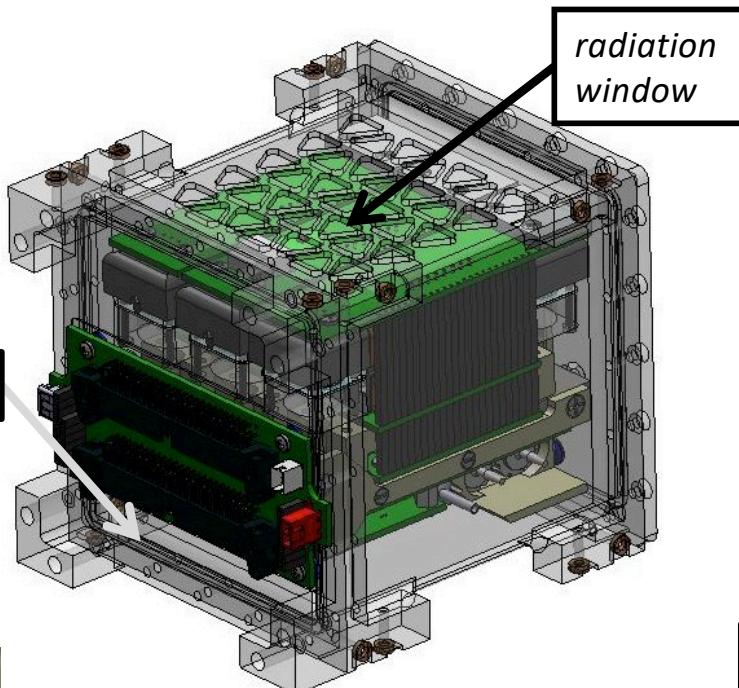




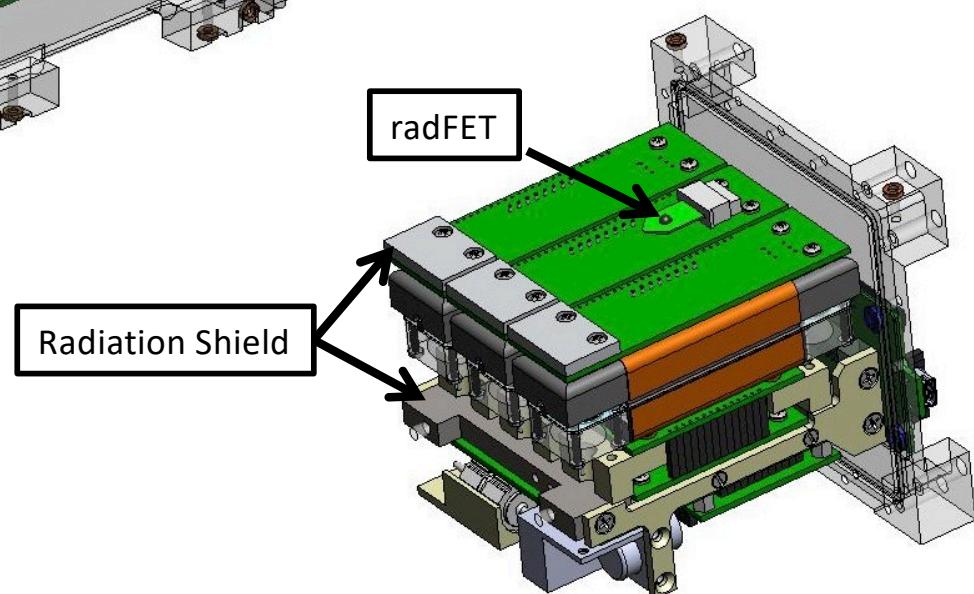
SESCO payload on O/OREOS

Biological / fluidic / optical / thermal cross-section





SESL Integration Summary

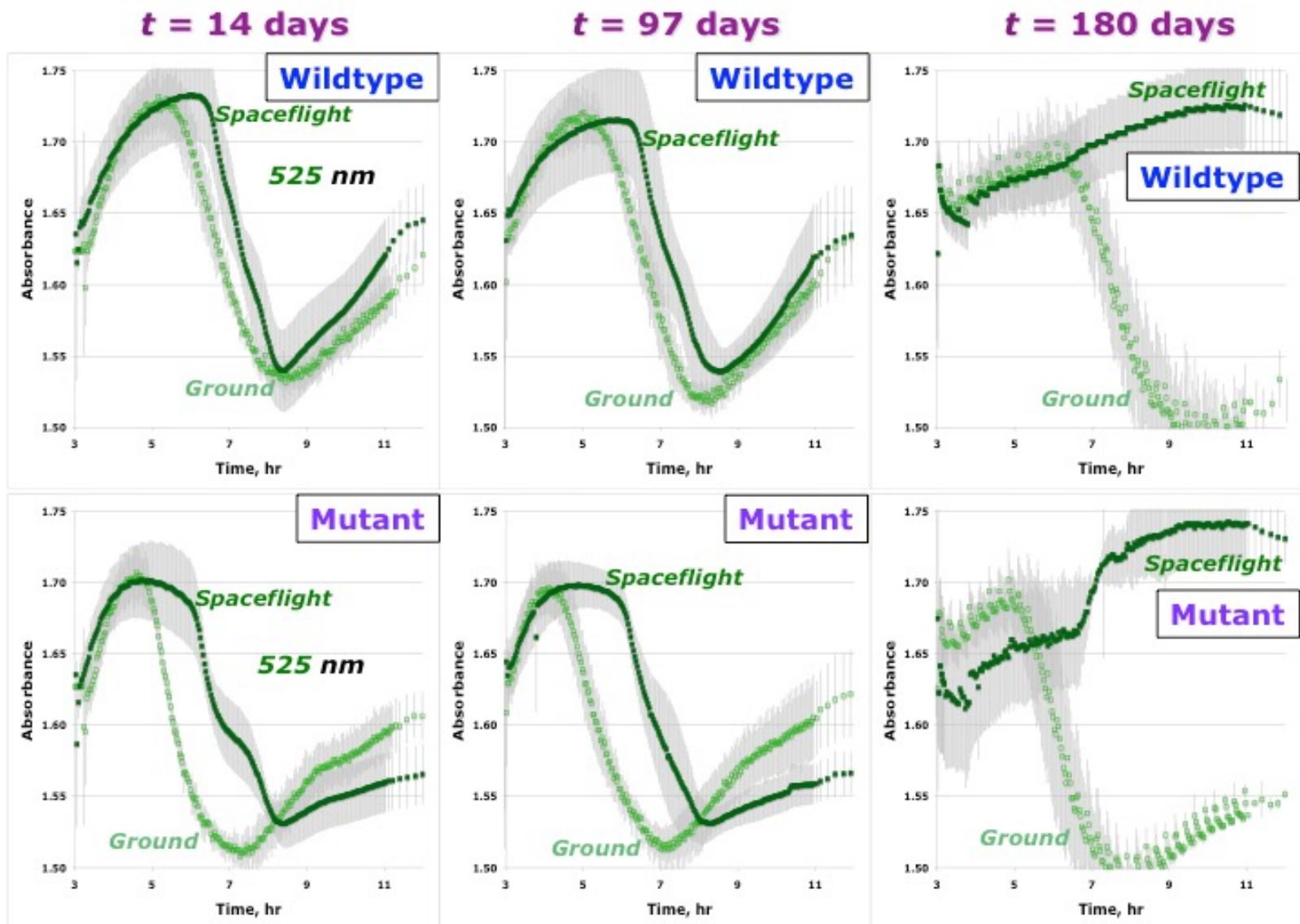




SESL
Spaceflight
Results:
Growth of
***B. subtilis* in**
space vs.
ground

WL Nicholson,
AJ Ricco, *et al*,
Astrobiology, 11,
951-958 (2011)

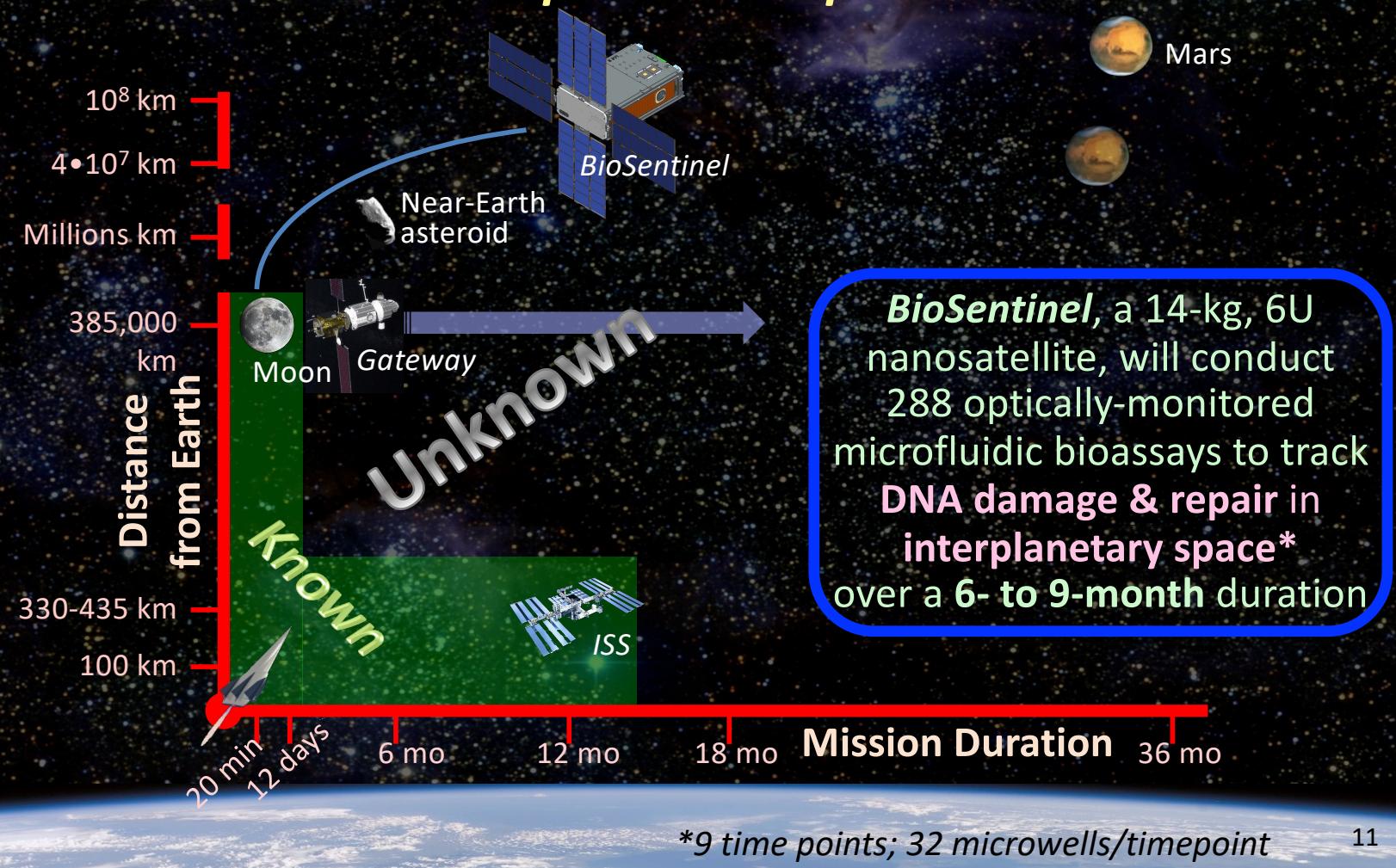
WL Nicholson
and AJ Ricco,
Life, 10, 1-14
(2020)





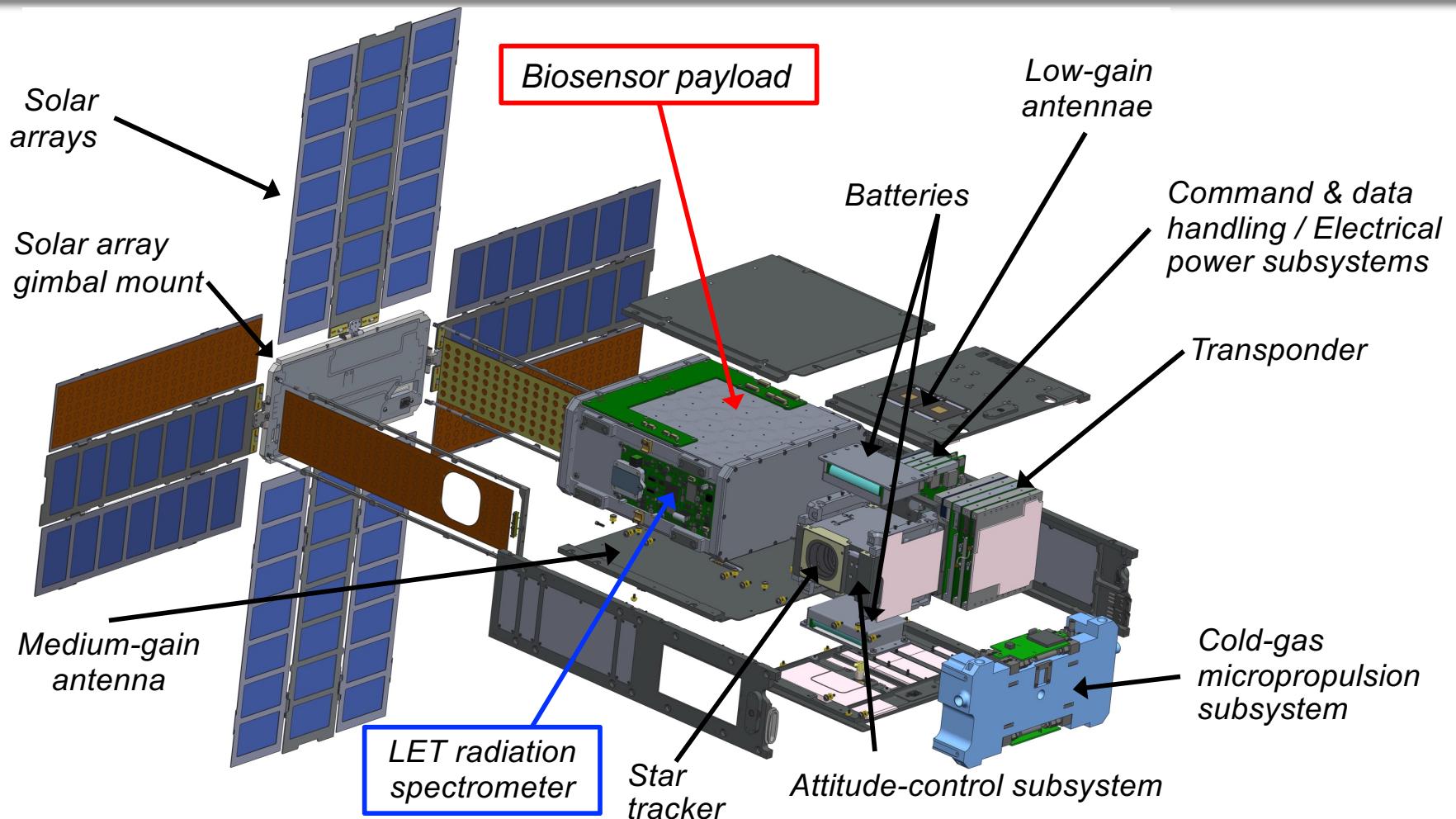
Biomapping our solar system: going beyond the known low-altitude / short-duration mission parameter space...

BioMapping:
Looking for
Life
Signatures
and testing
Terrestrial
Life
in many
locations





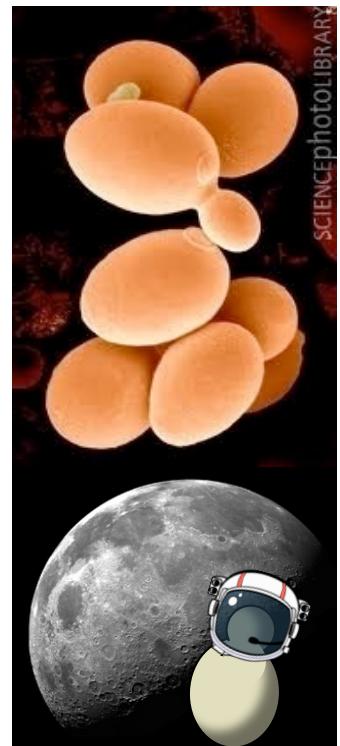
BioSentinel Overview: Taking Biofluidic Systems to Deep Space





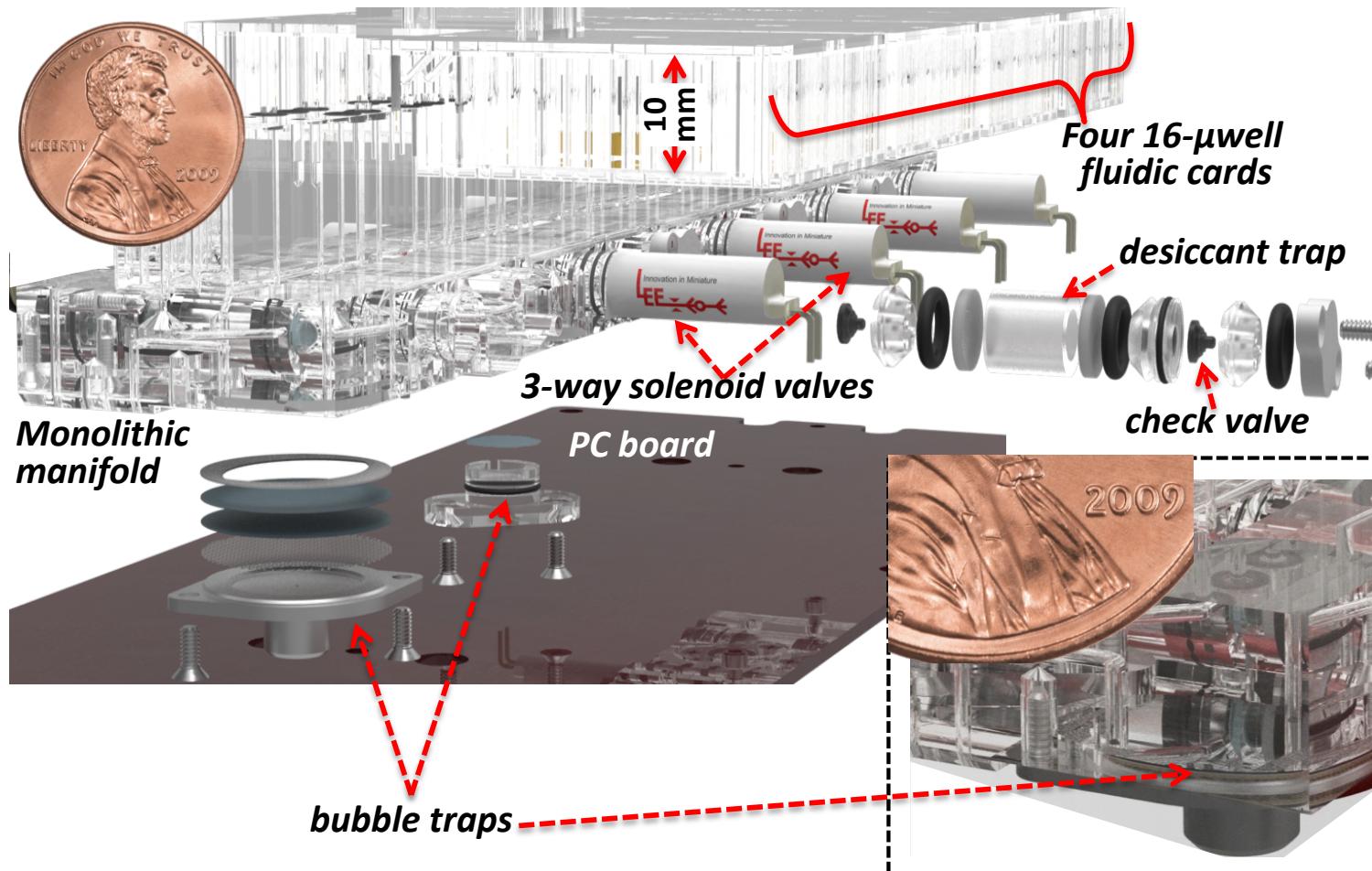
BioSentinel Science Mission: “*Canary in a Coal Mine*”

- **Quantify DNA damage from space radiation environment**
 - Deep space environment cannot be reproduced on Earth: *omnidirectional, continuous, low flux, variety of particle types*
 - Health risk for humans spending long durations beyond LEO
 - Radiation flux can spike 1000x during a solar particle event (SPE)
- **Yeast assay: microfluidic arrays monitor DNA damage**
 - Two strains of *S. cerevisiae*: 1 control (wild-type), 1 mutant
 - *engineered strain is sensitive to DNA damage, esp. double-strand breaks (DSBs)*
 - Wet and activate multiple banks of yeast in μ wells over mission duration
 - DNA damage impairs cell growth & division, esp. for *rad51 Δ* mutant
- **Correlate biological response with physical radiation measurements**
 - **Linear Energy Transfer** (LET) spectrometer bins and counts particle events by their LET
 - Total Ionizing Dose (TID): calculation of integrated deposited energy by LET system





Predecessor to Search-for-Life Fluidic Technologies: BioSentinel's Fluidic Processor, aka manifold

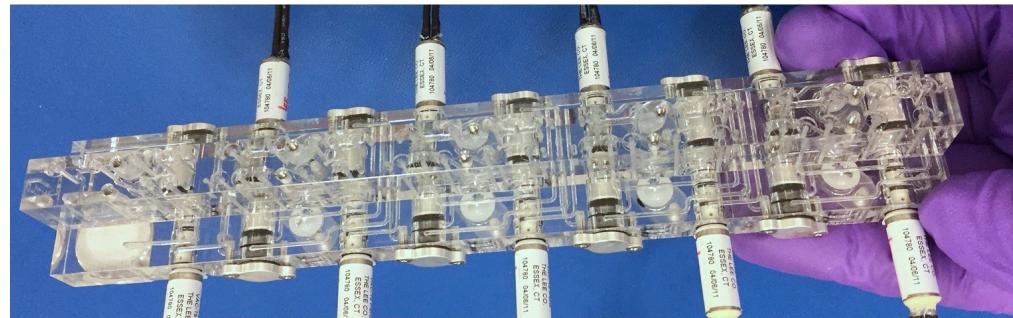


The fluidic analogy of multilevel printed-circuit boards

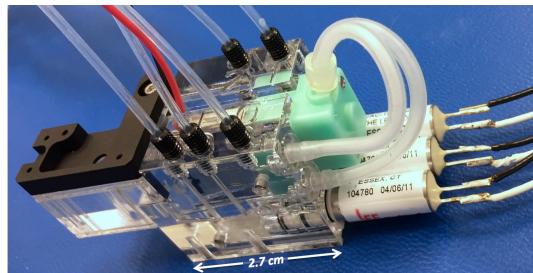


BioSentinel

Biofluidic Subsystem



9-fluidic-card manifold (144 wells) [1 of 2]

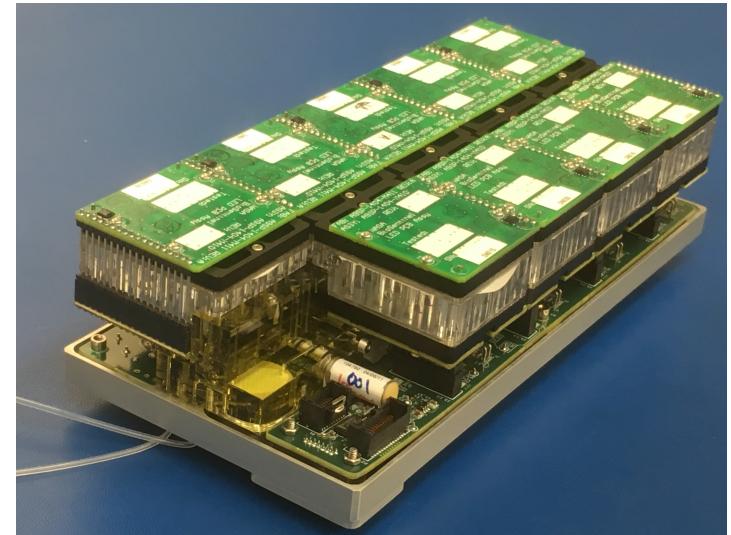


Reagent-and-pump manifold [1 of 2]

Tally of components:

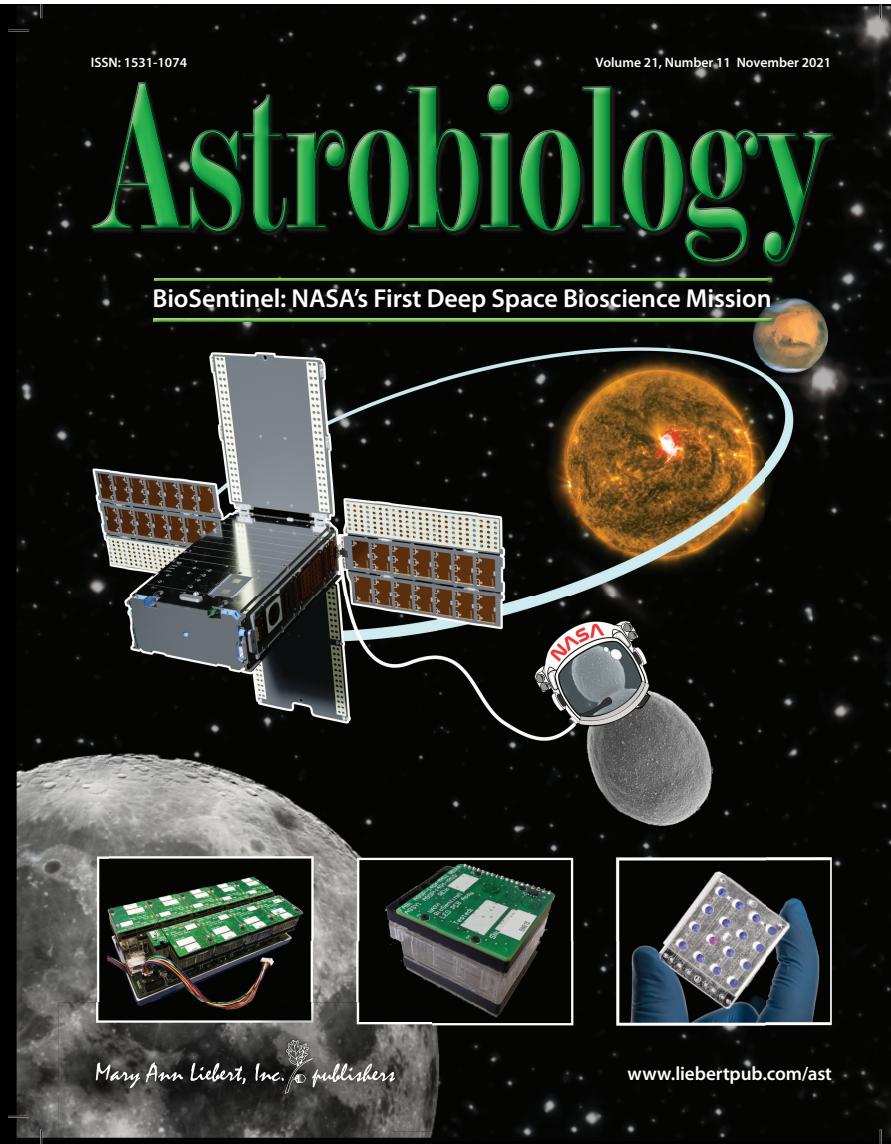
- 2 pumps, 2 main bubble traps
- 24 active valves, 38 check valves
- 16 fluidic cards with 16 small bubble traps, 16 desiccant traps, 288 wells total

9 fluidic cards integrated with measurement optics, thermal control, and fluidic manifold [1 of 2]



MR Padgen *et al.*,
Astrobiology, 21(5),
11 pp (2021).

AJ Ricco *et al.*,
*IEEE Aerospace
Elect Sys Mag*, 35,
18-24 (2020)

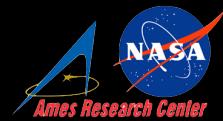




Ames Biological Small Satellite Payloads & Missions

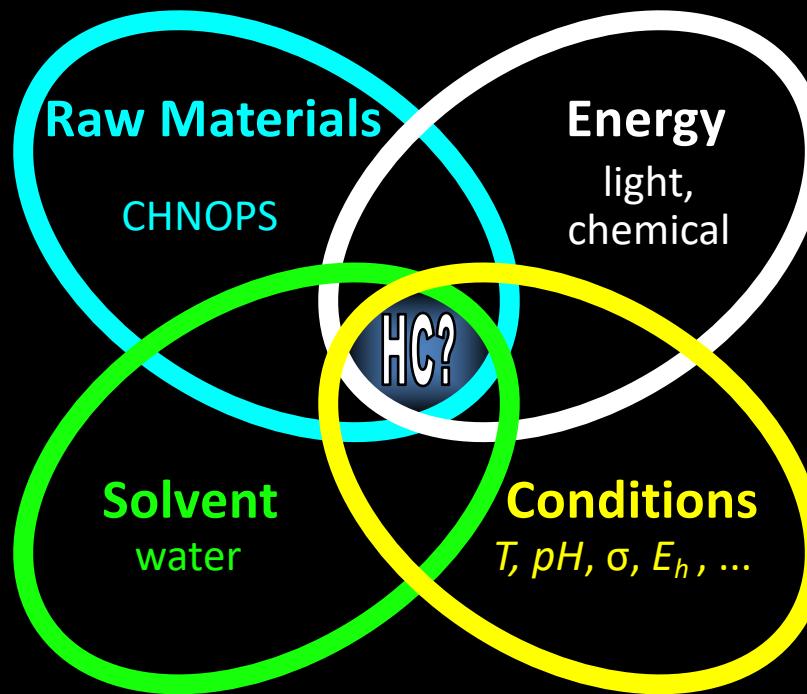
Mission* (format)	Description	Launch Date	Outcome
GeneSat-1 (3U)	<ul style="list-style-type: none"> 2U payload measured expression of GFP in <i>E. coli</i> and tracked microbe population via light scattering 1st NASA nanosatellite mission; 1st biological payload to fly in space in cubesat platform 	2006	Full mission success
PharmaSat (3U)	<ul style="list-style-type: none"> 2U payload measured antifungal drug dose response for <i>S. cerevisiae</i> using colorimetry to measure metabolic activity and population vs. time 1st NASA PI-led nanosatellite mission 	2009	Full mission success
O/OREOS (3U)	<ul style="list-style-type: none"> Two independent 1U astrobiology payloads measured (a) survival of <i>B. subtilis</i> to 6 months and (b) photo-degradation of biomarkers and bio-building blocks for > 1.5 years via UV-visible spectroscopy High-radiation high-inclination orbit; de-orbit mechanism; cells rehydrated in space 	2010	Full mission success, both payloads Spacecraft operable ~ 5 years in orbit
GraviSat (3U)	<ul style="list-style-type: none"> 2U payload integrating microcentrifuge to support microalgae (<i>C. vulgaris</i>, <i>D. bardawil</i>) in multiple wells with pulse-amplitude-modulated fluorescence & carbonate ion measurements 	N/A	Payload qual'd. to TRL 5
MisST (3U)	<ul style="list-style-type: none"> 2U payload integrating a multi-strain <i>C. elegans</i> fluidic habitat with a 2-color fluorescence microscope 	N/A	Payload qual'd. to TRL 6
SporeSat-1 SporeSat-2 (3U)	<ul style="list-style-type: none"> 2U payload measures gravitational response of <i>C. richardii</i> fern spores via Ca²⁺ ion channel response variable g in microgravity ambient using 50-mm pcentrifuges with 32 ion-selective [Ca²⁺] electrode pairs 	SporeSat-1: 2014 SporeSat-2: tbd	SporeSat-1: Successful spaceflight demo. of pcentrifuges & integral ion-selective electrodes
EcAMSat (6U)	<ul style="list-style-type: none"> 2U payload measured antibiotic resistance in microgravity vs. dose for uropathogenic <i>E. coli</i> 1st biological nanosat deployed from ISS; 1st Ames 6U biological nanosat 	2017	Full mission success
Eu:CROPIS / Powercells ("minisat")	<ul style="list-style-type: none"> Secondary payload, aboard DLR (German Aerospace Center) rotating 1-meter-diameter, 250-kg artificial-gravity satellite, to demonstrate synthetic biology-relevant processes 	2018	Success Further details TBD
BioSentinel (6U)	<ul style="list-style-type: none"> 4U payload to measure radiation-induced DNA damage in radiation-sensitive <i>S. cerevisiae</i> strain and correlate with physical radiation dosimetry and spectroscopy NASA Ames' 1st deep space nanosat.; 1st biology experiment beyond low Earth orbit since Apollo era 	2021(?)	TBD

The Search for Life & Habitability



Environmental Requirements to Sustain Life

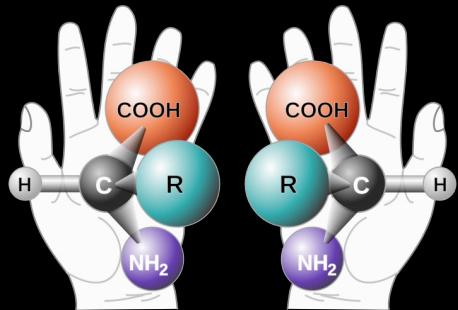
HC = HABITABLE CONDITIONS



Present habitability easier to assess than Past...

Searching for Life: What to Detect?

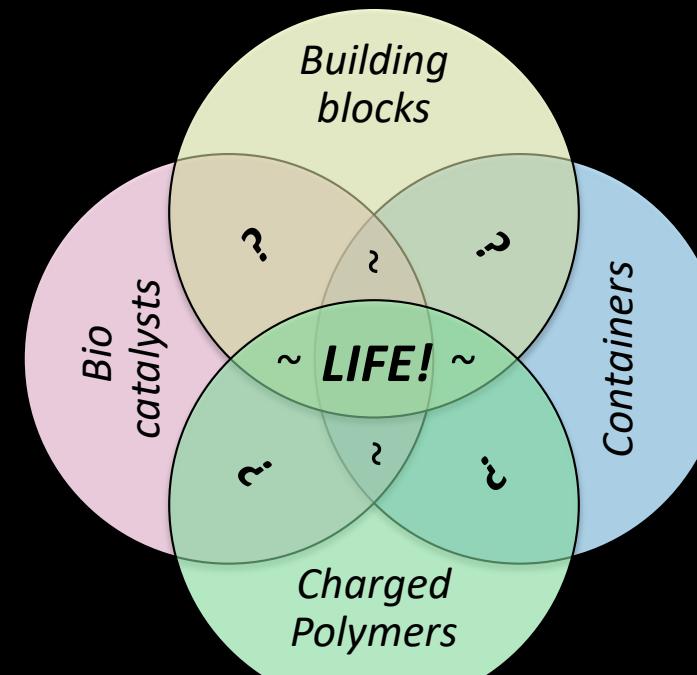
some aspects of life are likely to be universal...



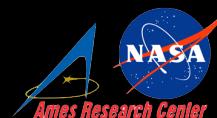
- Versatile chemical building blocks
- Complex multimeric biomolecules
- Containment structures
- Function-specific molecules

Arguably, all are required for life

- Combined, these indicators could provide conclusive evidence of life
- *Technologies needed to enable the search in an icy-moon environment?*

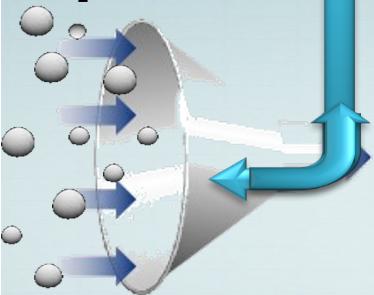


Targets, Rationale, and Instruments for Life Search



Measurement Target	Observed Parameter	Life Detection Rationale examples	Analytical Approach
Molecular building blocks	Chirality	Enantiomeric excess enables biochemistry <i>amino acids, saccharides</i>	<i>Capillary Electrophoresis</i> <i>Mass Spec</i>
Functional molecules	Catalysis	Biochemical processes; electron transfer <i>kinases; quinones</i>	<i>Electrochemical BioSensors</i> <i>Mass Spec</i>
Biogenic organic polymers	'Structural' polymers	Containers, energy, biochemistry <i>lipids</i>	<i>Mass spec</i> <i>Capillary Electrophoresis</i>
	'Information' polymers	Information storage and transfer <i>poly nucleic acids</i>	<i>Sequencing</i> <i>Mass Spec</i>
Containers	Morphology	Containers, structures, barriers <i>cells, membranes</i>	<i>Fluorescence Microscopy with staining</i>

Sample Input



Fluidics Processor

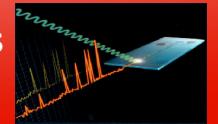
1. Deliver extraction solution
2. Retrieve sample with particles
3. Separate particles 4. Add dyes
5. Degas / de-bubble
6. Adjust ionic strength
7. Remove interfering ions
8. Adjust pH or solvent polarity
9. Adjust solvent polarity
10. Concentrate samples
11. Reconstitute standards/reagents
12. Provide calibration standards
13. Provide controls / blanks
14. Deliver particle-free aliquots

Instrument Suite Candidates

Fluorescence microscopy



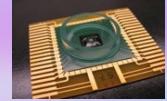
Microchip capillary electrophoresis with laser-induced fluorescence



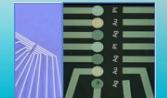
Mass spectrometry w/ electrospray, GC, or (MA)LDI "front end"



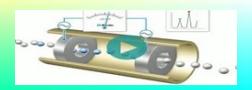
Electrochemical biosensors



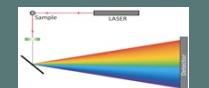
Ion-selective electrodes [Habitability, energy]



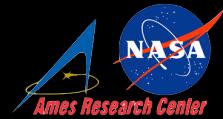
Ion chromatography [Habitability]



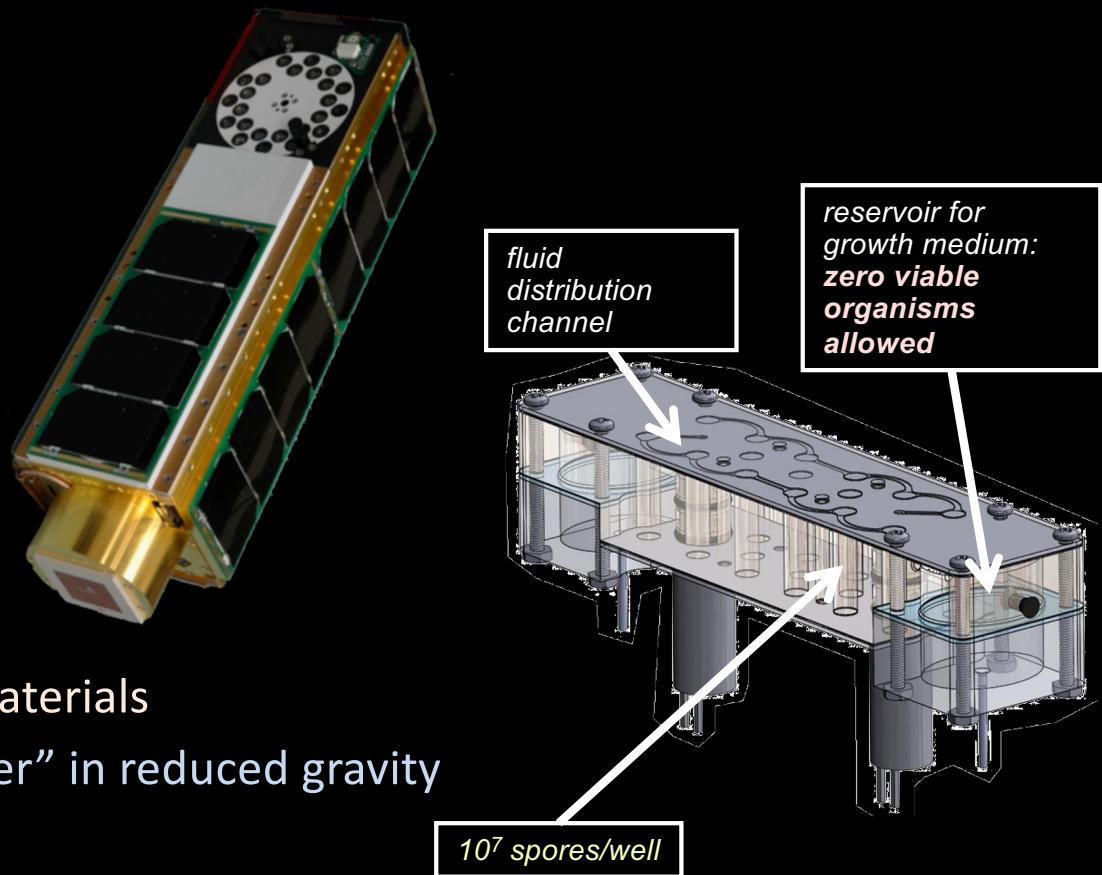
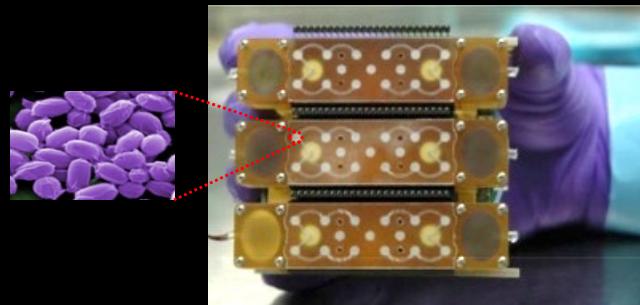
Raman spectroscopy



Past Biological Space Missions provide Enabling Technologies



O/OREOS



Relevant payload technologies

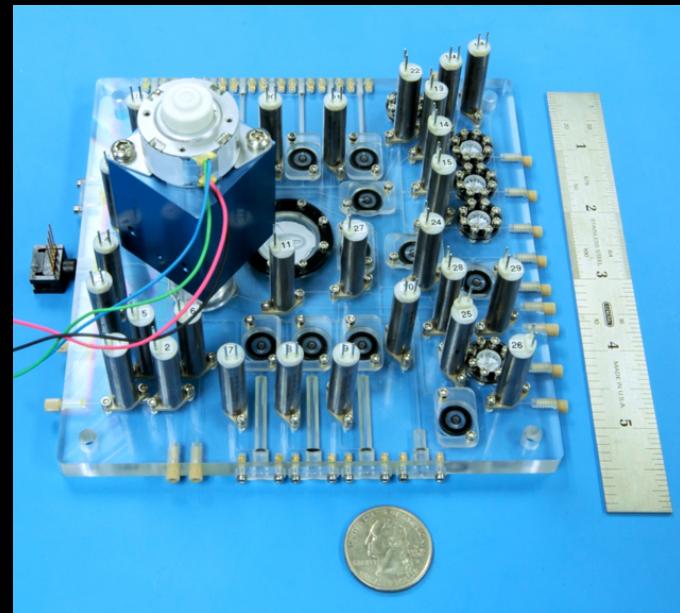
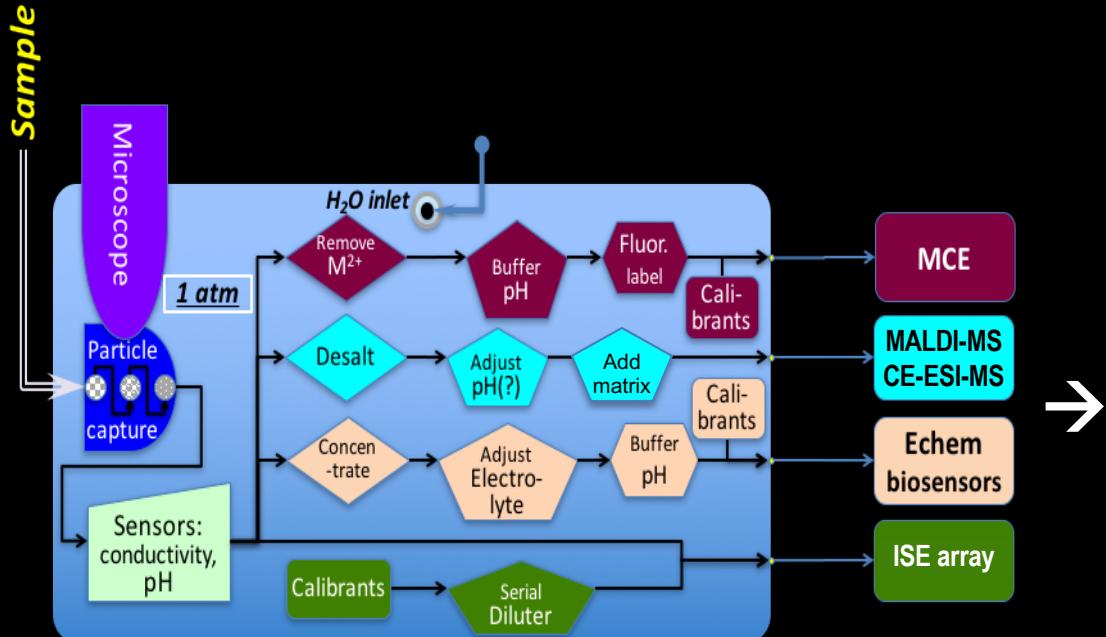
- Handling biological specimens in space
- Perfect sterility (flight-proven)
- Ultra-low organic contamination
- Biocompatible, low-leach-and-offgas materials
- Fly dry, then wet-out fluidics “much later” in reduced gravity
- Manipulate nL – μ L volumes
- Functional in high-radiation environment



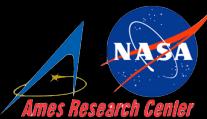
Integrated Life Detection Payloads

Sample Processor for Life on Icy Worlds (*SPLIce*):

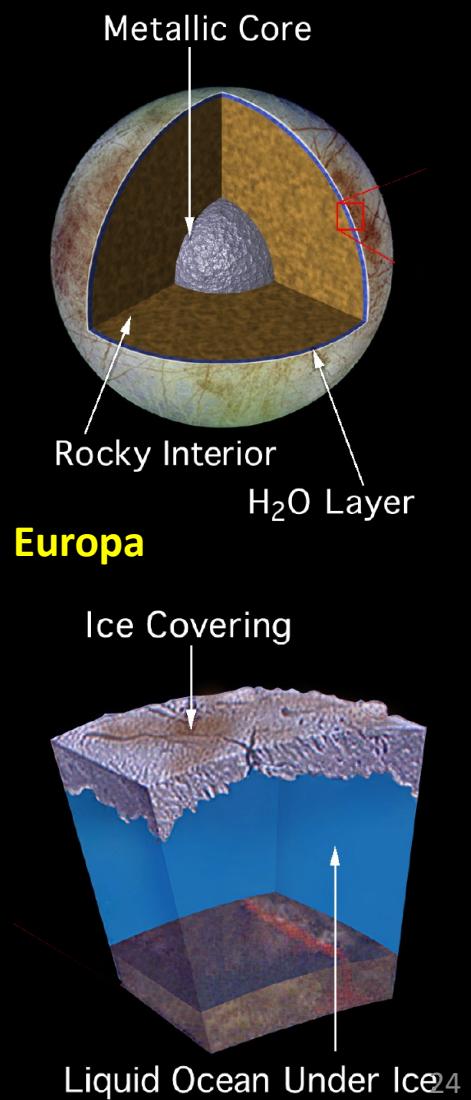
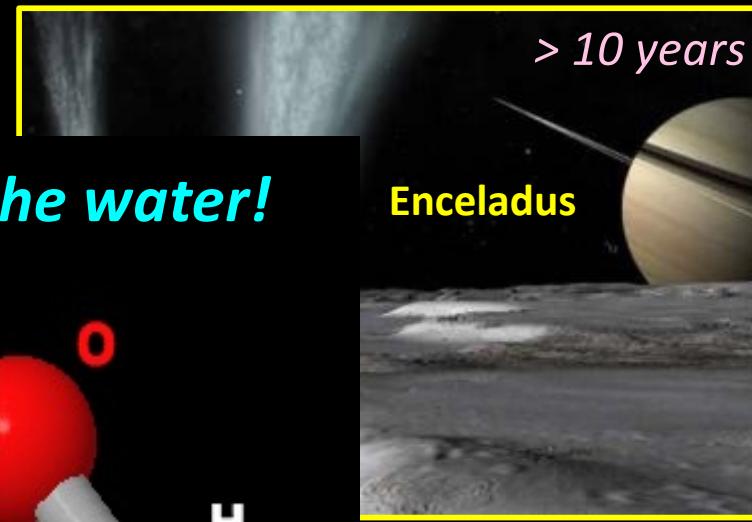
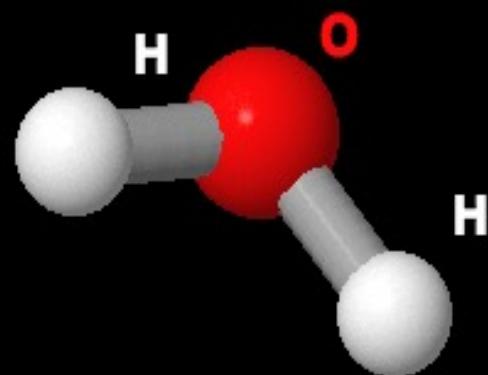
- *Technology development for Enceladus & Europa life search*
- Partners: APL (Johns Hopkins U.), NASA/JPL, NASA/Goddard, Tufts University



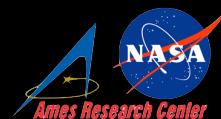
SPLIce Monolithic Manifold



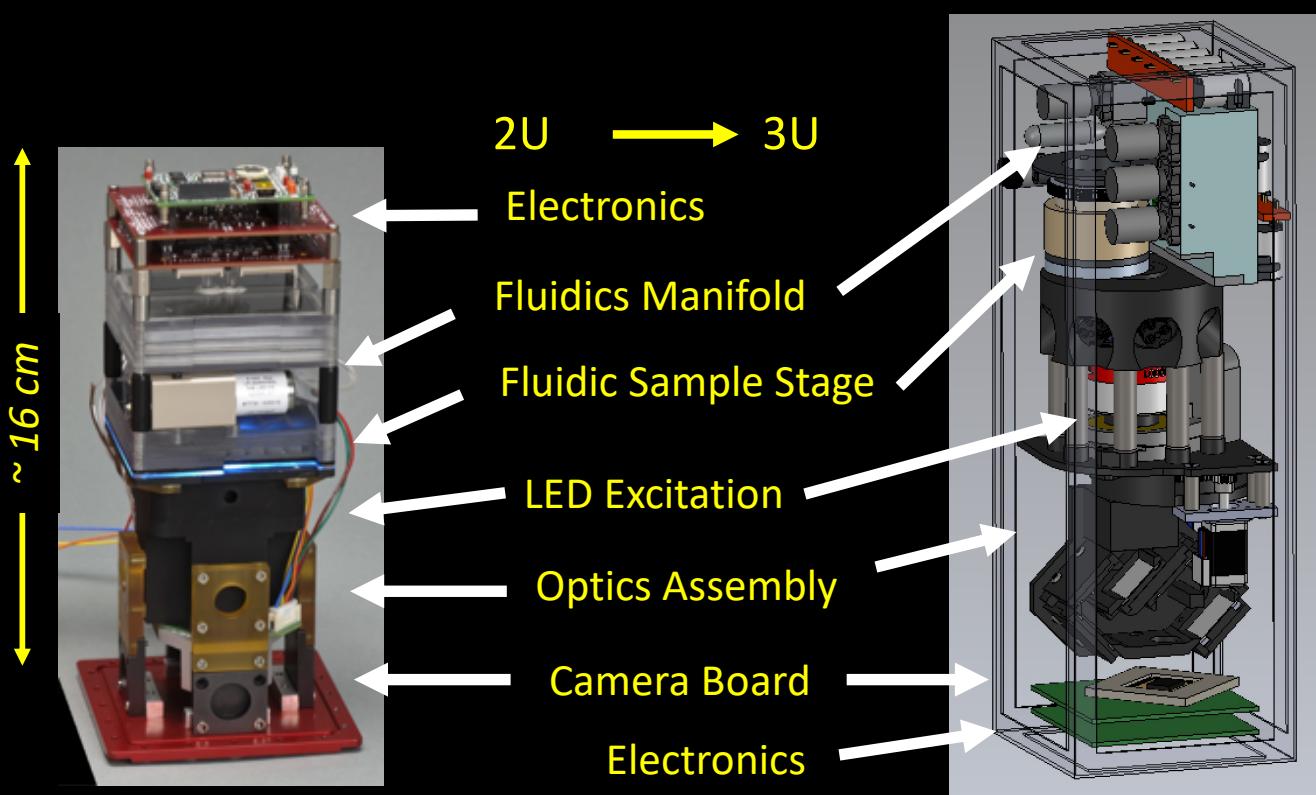
Life-Search Exploration Targets



ELM: Europa Luminescence Microscope



Based on ARC's nanosatellite 2U **fluorescence & fluidics** imager for *C. elegans*



P. boydii ~ 1 μm Wide



Cell Mask – Membrane Lipids

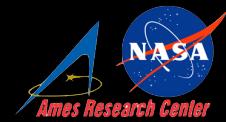


Alexa Fluor 405 - Membrane Proteins

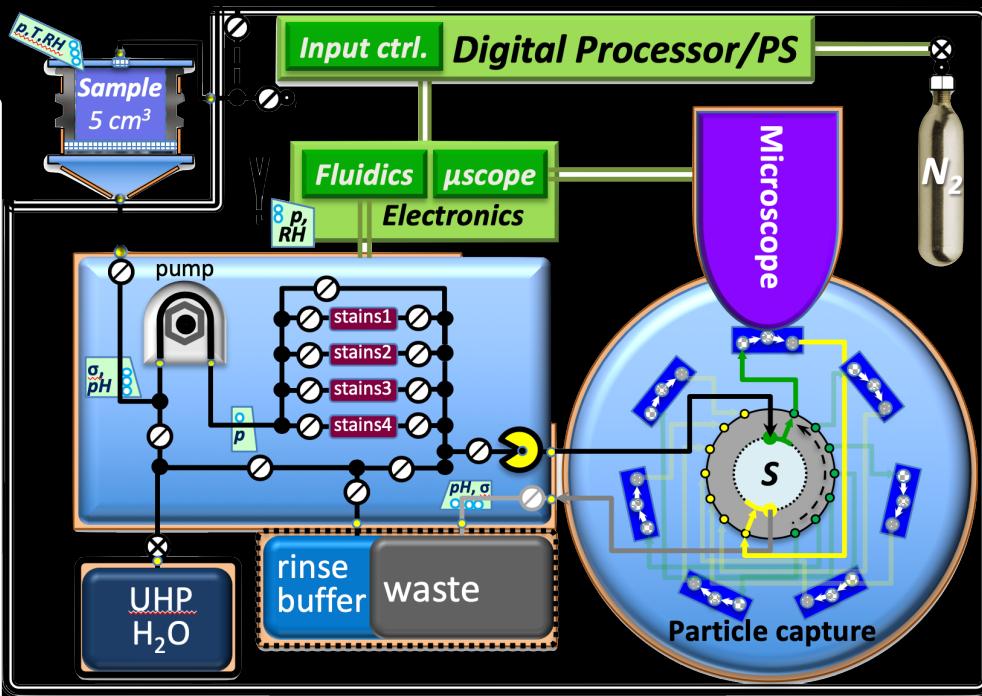


SYTO9 – Nucleic Acids

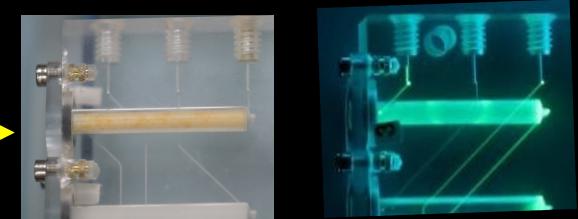
Europa Luminescence Microscope – Block Diagram & Fluidics Breadboard



MisST Imager: 2U
Nanosat payload



Fluorescent stains: Porous-polymer supports
for anhydrous storage, aqueous rehydration



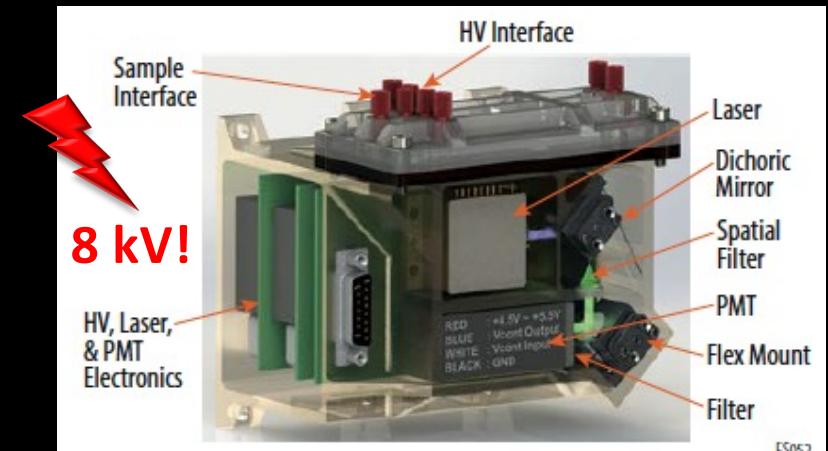
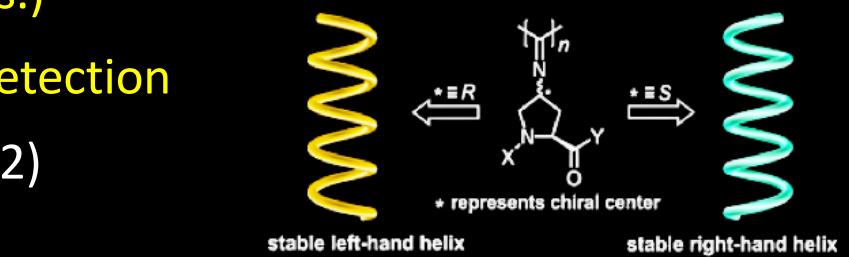
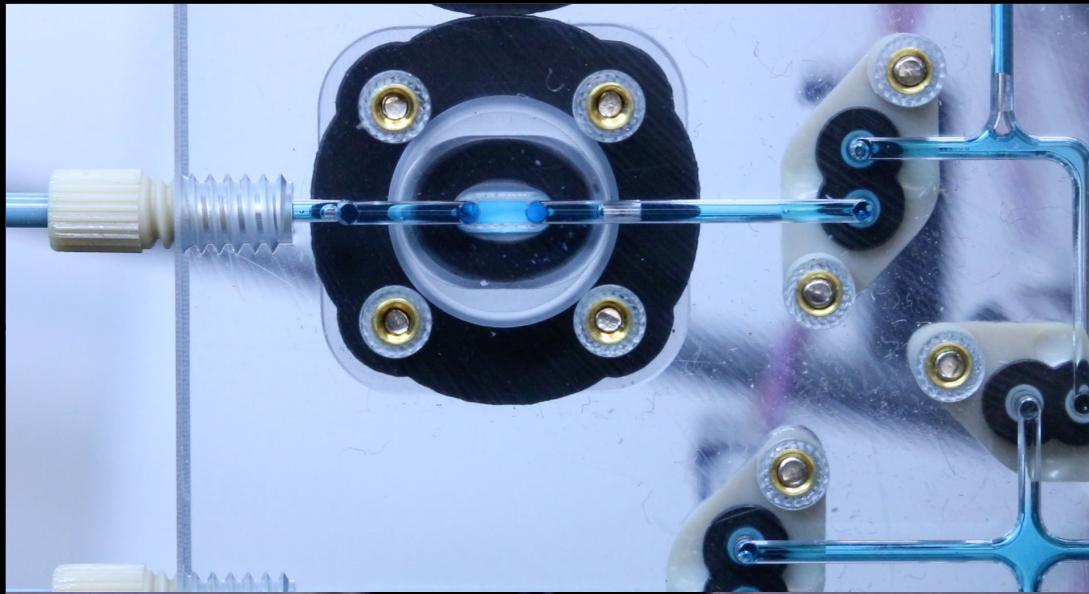
Microchip Capillary Electrophoresis (JPL, ARC)

Chiral separations, e.g. amino acids (JPL: field demos.)

- Ultrasensitive laser-induced-fluorescence detection

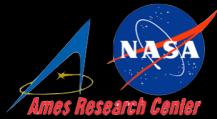
ARC sample-processing “front end” (COLDTech/ICEE2)

- Combination bubble trap / air-gap generator



MICA: Microfluidic Icy-world Chemical Analyzer

Fluidically Integrated Habitability Assessment for Icy Worlds

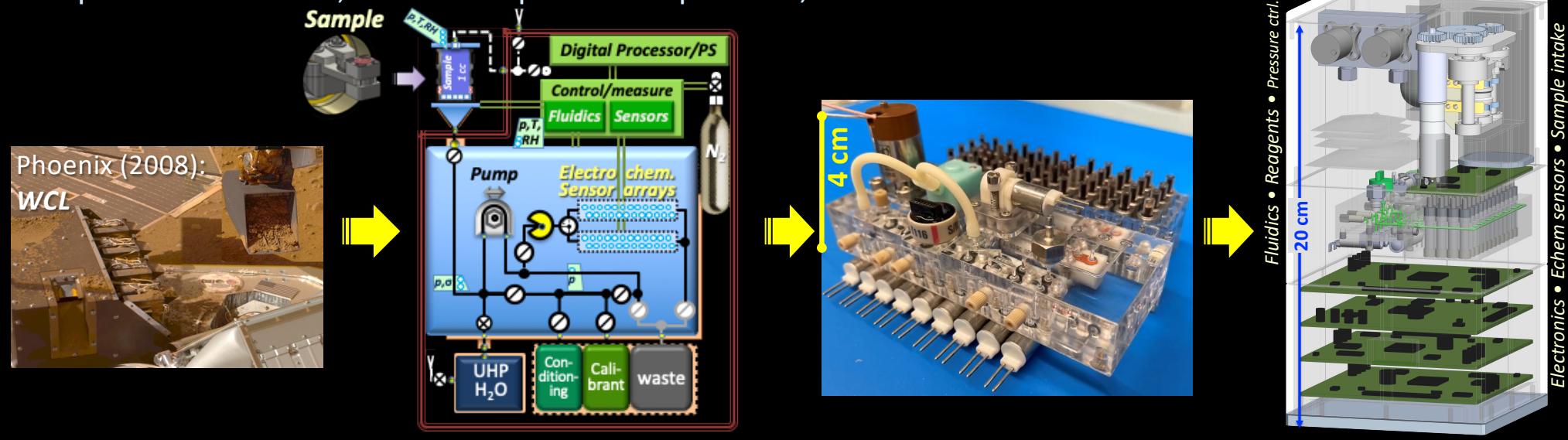


Heritage: Phoenix Wet Chemistry Lab (WCL) + COLDTech, ICEE-2 projects

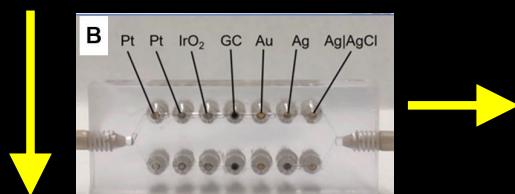
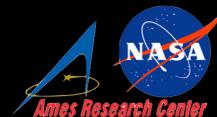
Lead: ARC **Partners:** JPL, Tufts, MIT, U. of Alberta, Honeybee Robotics

Key Measurements: pH, conductivity, ions, gases, ROS: Li⁺, Na⁺, K⁺, Mg²⁺, Ca²⁺, Cl⁻, CO₃²⁻, NH₄⁺, NO₃⁻, ClO₄⁻, ClO₃⁻, PO₄³⁻, SO₄²⁻, SO₃²⁻; O₂, SO₂, O₂⁻, H₂O₂; solution energetics: discrete & average of redox-active species (E_h)

Fluidic Functions : Receive & melt icy samples; prepare/deliver conditioning, blank, & calibrant solutions at multiple concentrations; control temperature & pressure; execute & store measurements

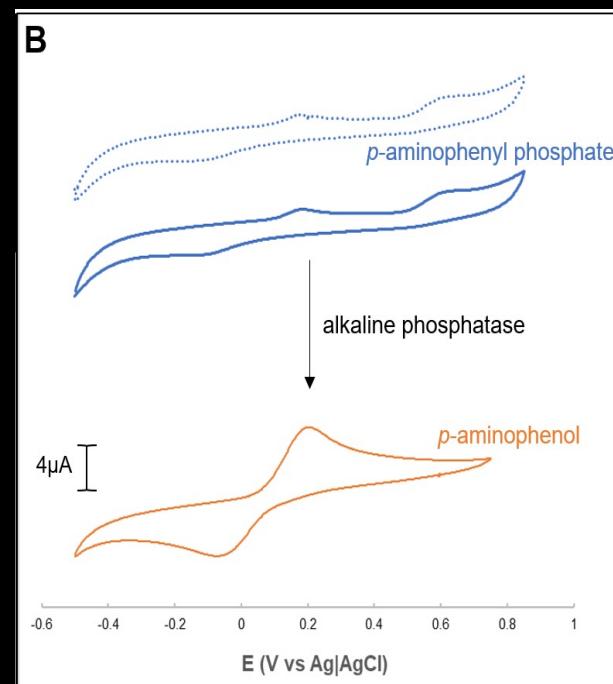


Electrochemical Detection of Catalysts as Signatures of Life

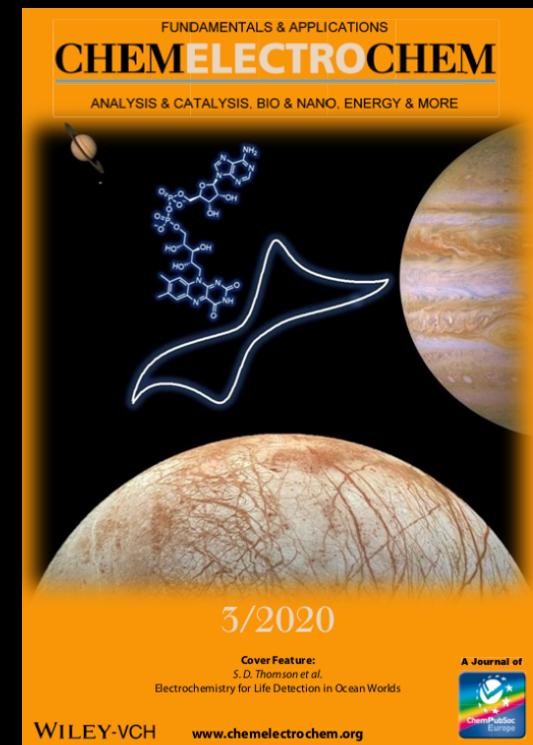


MICA 2021
Ames, JPL, Tufts U., MIT

- Leverages Phoenix Wet Chemistry Lab technology
- Example targets: kinases, phosphatases, proteases
- Same instrument characterizes habitability & energetics



SD Thomson *et al.*,
ChemElectroChem 2020



WILEY-VCH

www.chemelectrochem.org



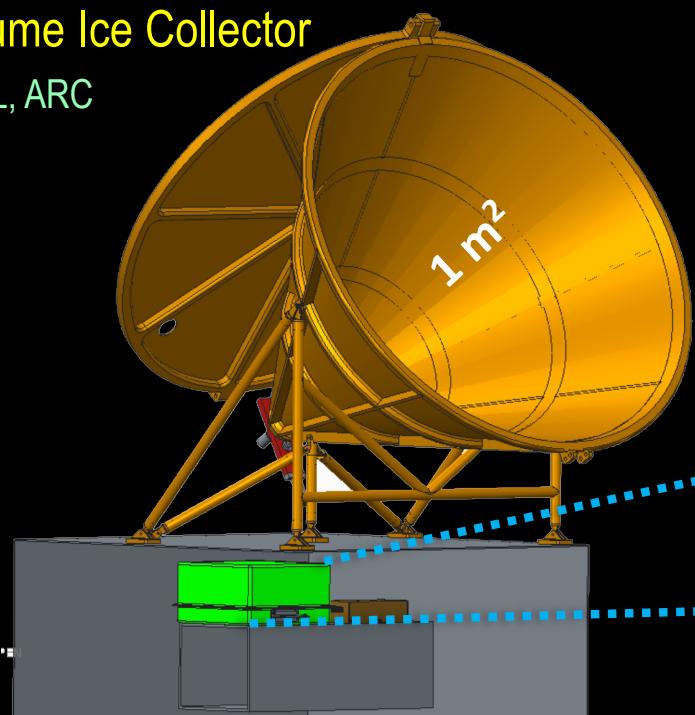
ELSAH: Enceladus Life Signatures and Habitability

New Frontiers 4

µCAFE: micro Chemical Analyzer of Fluids for Exobiology

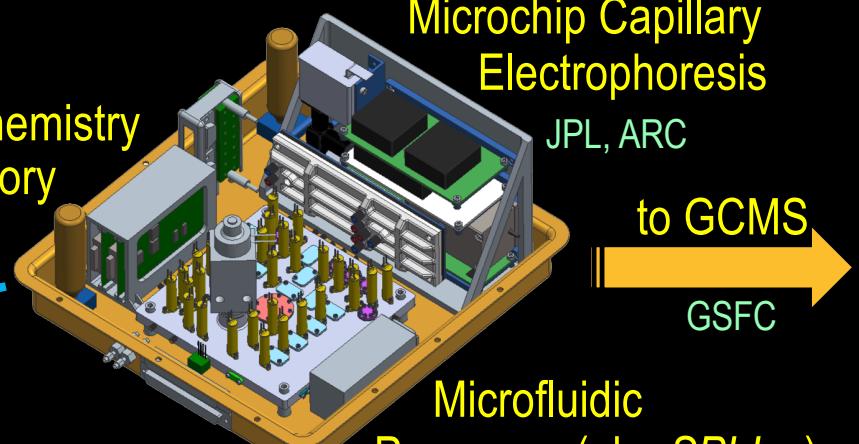
Plume Ice Collector

APL, ARC



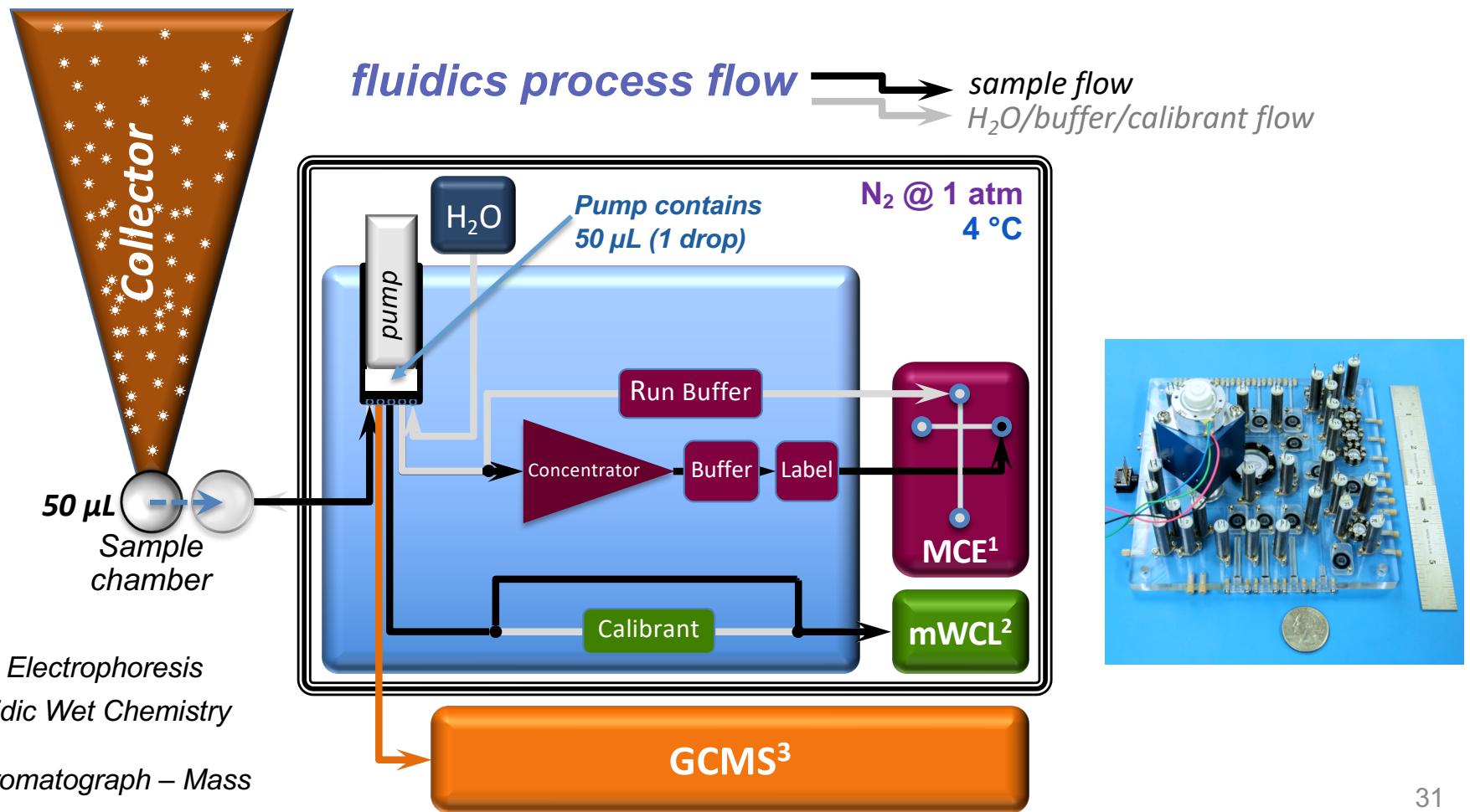
Tufts, ARC, JPL, MIT

micro-Wet Chemistry
Laboratory



Microfluidic Processor (aka SPL/ice)
ARC & partners

Sample Processor for Life on Icy worlds: “SPLIce” as the core of a Life-Search System for Enceladus





EcAMSat above Earth

Conclusions

ELSAH below Enceladus



- **HERITAGE of astro- & space-biology experiments in low-Earth orbit is a major enabler for interplanetary astro/biological missions, including the search for life on icy moons**
 - Full autonomy, flying dry, filling fluidic systems in μ -gravity, managing bubbles, ...
 - Long-term materials biocompatibility, perfect sterility, contamination control
 - Radiation-tolerant design: *O/OREOS* functional to 5 years (15x ISS radn. levels); *BioSentinel* design for 2 yr in deep space, fluidics radn. sterilized at 3.5 Mrad (35 kGy)
 - High-heritage sensors, fluidic components & approach; high-efficiency / uniformity thermal control, managing pressure and condensation ...



Thanks!

O/OREOS: Pascale Ehrenfreund, John Hines, Dave Squires, Chris Kitts, Charlie Friedericks, Dave Landis, Elwood Agasid, Matthew Piccini, Chris Beasley, Nathan Bramall, Gio Minelli, Greg Defouw, Laura Bica, Katie Bryson, Roland Burton, Julie Chittenden, Amanda Cook, Millan Diaz-Aguado, Shak Ghassemieh, Mike Henschke, Ed Luzzi, Diana Ly, Nghia Mai, Rocco Mancinelli, Andy Mattioda, Mike McIntyre, Mike Neumann, Wayne Nicholson, Macarena Parra, Richard Quinn, Mike Rasay, Bob Ricks, Orlando Santos, Aaron Schooley, Eric Stackpole, Linda Timucin, Bruce Yost, Anthony Young

NASA/Ames, Santa Clara U., George Washington U., Draper Lab, SETI, U. Florida/KSC

BioSentinel: Bob Hanel, Dawn McIntosh, Brian Lewis, Charlie Friedericks, Sharmila Bhattacharya, Matt Dortenzio, James Chartres, Ben Bradley, Zion Young, Tom Luzod, Chris Storment, Macarena Parra, Sergio Santa Maria, Diana Marina, Lauren Liddell, Sofia Tieze, Abe Rademacher, Josh Benton, Terry Lusby, Mike Padgen, Travis Boone, Ming Tan, Lance Goddard, Aliyeh Mousavi, Diana Gentry, Aaron Schooley, Matthew Sorgenfrei, Matthew Nehrenz, Vanessa Kuroda, Ben Klamm, Craig Pires, Shang Wu, Doug Forman, Hugo Sanchez, Elwood Agasid, Tore Straume, Bobbie Gail Swan, Scott Wheeler, Susan Gavalas, Greg Nelson, Troy Harkness

NASA/Ames, NASA/JSC-Radworks, Loma Linda U. Med. Ctr., U. Saskatchewan

ARC Search-for-Life Technologies Team: Richard Quinn, Mary Beth Wilhelm, Justin Blaich, Travis Boone, Nathan Bramall, Kathryn Bywaters, Matthew Chin, Tori Chinn, Chris Espinoza, Josh Forgione, Lauren Friend, Nelson Gaspard, Trinh Hoac, Erin Kelly, Dayne Kemp, Greg Kintz, Anthony Lee, Tom McClure, Griffin McCutcheon, Connor Nelson, Abraham Rademacher, Leslie Radosevich, Jared Shimada, Ming Tan, Linda Timucin, Jonathan Wang, and Pete Zell

NASA/Ames

\$ **GeneSat: NASA Fundamental Space Biology, ESMD (now ~ SLPSRA/HEOMD)**

\$ **O/OREOS: NASA Astrobiology Small Payloads Program, SMD**

\$ **BioSentinel: NASA Advanced Exploration Systems, HEOMD**

\$ **Search for Life Tech.: NASA COLDTech and ICETE2 Programs, SMD**